DATA NOTE

A curated transcriptome dataset collection to investigate the blood transcriptional response to viral respiratory tract infection and vaccination. [version 1; peer review: 2 approved]

Salim Bougarn, Sabri Boughorbel, Damien Chaussabel, Nico Marr

Systems Biology and Immunology Department, Sidra Medicine, Doha, Qatar

Abstract

The human immune defense mechanisms and factors associated with good versus poor health outcomes following viral respiratory tract infections (VRTI), as well as correlates of protection following vaccination against respiratory viruses, remain incompletely understood. To shed further light into these mechanisms, a number of systems-scale studies have been conducted to measure transcriptional changes in blood leukocytes of either naturally or experimentally infected individuals, or in individual's post-vaccination. Here we are making available a public repository, for research investigators for interpretation, a collection of transcriptome datasets obtained from human whole blood and peripheral blood mononuclear cells (PBMC) to investigate the transcriptional responses following viral respiratory tract infection or vaccination against respiratory viruses. In total, Thirty one datasets, associated to viral respiratory tract infections and their related vaccination studies, were identified and retrieved from the NCBI Gene Expression Omnibus (GEO) and loaded in a custom web application designed for interactive query and visualization of integrated large-scale data. Quality control checks, using relevant biological markers, were performed. Multiple sample groupings and rank lists were created to facilitate dataset query and interpretation. Via this interface, users can generate web links to customized graphical views, which may be subsequently inserted into manuscripts to report novel findings. The GXB tool enables browsing of a single gene across projects, providing new perspectives on the role of a given molecule across biological systems in the diagnostic and prognostic following VRTI but also in identifying new correlates of protection. This dataset collection is available at: http://vri1.gxbsidra.org/dm3/geneBrowser/list.

Keywords
Transcriptomics, Bioinformatics, Software, Viral respiratory infection, Influenza viruses, Respiratory syncytial viruses (RSV), Rhinoviruses, Whole Blood, PBMC.

Open Peer Review

Approval Status  ✔  ✔

version 1
13 Mar 2019
view  view

1. Carlos Alberto Moreira-Filho,
University of São Paulo (USP), São Paulo, Brazil

2. Benjamin M Tang, Nepean Hospital,
Kingswood, Australia

Any reports and responses or comments on the article can be found at the end of the article.
**Corresponding author:** Salim Bougarn (sbougarn@sidra.org)

**Author roles:** Bougarn S: Conceptualization, Data Curation, Formal Analysis, Investigation, Methodology, Project Administration, Resources, Software, Validation, Visualization, Writing – Original Draft Preparation, Writing – Review & Editing; Bouhhorbel S: Data Curation, Resources, Software, Writing – Review & Editing; Chaussabel D: Conceptualization, Funding Acquisition, Project Administration, Supervision, Writing – Review & Editing; Marr N: Conceptualization, Project Administration, Supervision, Writing – Review & Editing

**Competing interests:** No competing interests were disclosed.

**Grant information:** All the authors listed on this publication received support from the Qatar Foundation. Support for this project was provided by the Qatar National Research Fund [NPRP10-0205-170348]. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Copyright:** © 2019 Bougarn S et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

**How to cite this article:** Bougarn S, Bouhhorbel S, Chaussabel D and Marr N. A curated transcriptome dataset collection to investigate the blood transcriptional response to viral respiratory tract infection and vaccination. [version 1; peer review: 2 approved] F1000Research 2019, 8:284 [https://doi.org/10.12688/f1000research.18533.1](https://doi.org/10.12688/f1000research.18533.1)

**First published:** 13 Mar 2019, 8:284 [https://doi.org/10.12688/f1000research.18533.1](https://doi.org/10.12688/f1000research.18533.1)
Introduction

Viral respiratory tract infections (VRTI) are responsible for the majority of hospitalizations among infants and the elderly. They are caused mainly by a heterogeneous group of viruses, including rhinoviruses, influenza viruses, parainfluenza viruses, respiratory syncytial virus (RSV), enteroviruses, coronaviruses, and certain strains of adenovirus\(^2\). Few antiviral therapies are currently approved and routinely used for VRTI. Most of these are specific inhibitors of influenza viruses\(^3\). Moreover, for most respiratory viruses, there is no licensed vaccine available\(^4\), with the exception of flu vaccines for which protection generally lasts only one flu season. Consequently, clinical management of individuals with VRTI is mostly restricted to supportive care\(^5\).

As clinical symptoms are often overlapping and are not specific for any of the viral species, it is difficult to establish a clinical diagnosis without laboratory testing\(^1\). Furthermore, clinical manifestations of VRTI are highly variable, ranging from asymptomatic infections or illness with mild symptoms (a common cold) to clinically severe disease with life-threatening complications, such as respiratory failure and in some cases may have a fatal outcome\(^6\). Infants, the elderly and patients with chronic lung or heart diseases in particular are at high risk\(^7\).

Thus, there is an evident need to better understand the molecular mechanisms underlying the disease pathogenesis, progression as well as severity of, and immunity against, VRTI among humans\(^8\). In this context, different large scale gene expression studies have been conducted using whole blood or peripheral blood mononuclear cells (PBMCs), to assess the human immune response to natural\(^9\) and experimental viral respiratory infections\(^10\), in particular, to influenza and RSV infections, and also to vaccination\(^11,12\).

Here, we make available, through an interactive web application, a curated collection of datasets that were obtained from pediatric and adult patients with natural VRTI, volunteers with experimental exposition to respiratory viruses and also vaccinated volunteers. Transcriptomics datasets were obtained from whole blood and PBMCs.

A total of 31 datasets were retrieved and selected from the NCBI Gene Expression Omnibus (GEO), a public repository of transcriptome profiles. The identified datasets are particularly relevant to our interest in understanding the pathobiology of VRTI and vaccination. As described in recent publications\(^13,14\), these datasets were loaded into a custom interactive web application, the Gene Expression Browser (GXB), which enables easy access to large datasets and interactive visualization of our dataset collection related to VRTI and vaccination against respiratory viruses. It also provides access to demographic and clinical information. Importantly, the user can customize data plots by adding multiple layers of parameters (e.g. age, gender, sample type, type of infection, type of vaccine, sample collection time), modify the sample ordering and genes, and generate links (mini URL) that can be shared via e-mail or used in publications. Therefore, we are providing here a resource enabling browsing of datasets relevant to blood transcriptional responses to VRTI and vaccination that offers a unique opportunity to identify host genes and their regulation that may be of diagnostic and/or prognostic value, or that may be tested as novel correlates of protection in subsequent studies. For example, a comparative approach of the transcriptional response signatures between experimentally infected and vaccinated individuals could be used to identify common mechanisms that define the poor health outcomes versus strong protection. The ability to pool, compare and analyze the immune responses to different infections and vaccines, in different individuals and at various age, offers a unique opportunity for a better understanding of the pathophysiology of VRTI.

Methods

A total of 120 datasets, potentially relevant to human immune responses to VRTI and vaccination, were identified in GEO using the following search query:


Most of retrieved datasets were generated from human blood and human PBMC, using Illumina or Affymetrix commercial platforms or RNA-sequencing. All the entries that were returned with this query were manually curated. The process involved reading all the descriptions available of the datasets, the study design and the GEO-linked article in pubmed. Finally, only studies using human whole blood and human PBMCs, associated with natural or experimental VRTI, or vaccination against VRTI, were retained for our dataset collection. For the retained datasets, if the platform used to generate the transcriptome profiles was not supported by GXB or if from an in vitro study, they were excluded from our dataset collection. Based on these criteria, 31 datasets were retained. These include datasets that were generated from whole blood or PBMCs of individuals who were either naturally (12) or experimentally infected (3) (with influenza viruses, RSV, Rhinovirus, Rotavirus) as well as from healthy, uninfected (age-matched) volunteers. The remaining 16 datasets were generated from whole blood or PBMCs of individuals who had received flu vaccines (Figure 1). The datasets that comprise our collection are listed in Table 1.

Once the final selection had been made, each dataset was downloaded from GEO by using the SOFT file format. Then, the datasets were uploaded on the Gene Expression Browser (GXB), an interactive web application hosted on the Amazon Web
Services cloud. Information about samples and study design were also uploaded. The available samples were put into groups based on relevant study variables and genes were ranked according to the different groups comparisons. A detailed description of the GXB software tool is available from recent publications. This software interface allows user to easily navigate and filter the dataset collection. A web tutorial can be easily accessed online. Annotation and functionality of the web software interface were described previously by our group, and is reproduced here so that readers can use this article as a standalone resource.

Briefly, datasets of interest can be quickly identified either by filtering on criteria from pre-defined sections on the left or by entering a query term in the search box at the top of the dataset navigation page. Clicking on one of the studies listed in the dataset navigation page opens a viewer designed to provide interactive browsing and graphic representations of large-scale data in an interpretable format. This interface is designed to present ranked gene lists and display expression results graphically in a context-rich environment. Selecting a gene from the rank ordered list on the left of the data-viewing interface will display its expression values graphically in the screen’s central panel. Directly above the graphical display drop down menus give users the ability: a) To change how the gene list is ranked - this allows the user to change the method used to rank the genes, or to only include genes that are selected for specific biological interest; b) To change sample grouping (Group Set button) - in some datasets, a user can switch between groups based on cell type to groups based on disease type, for example; c) To sort individual samples within a group based on associated categorical or continuous variables (e.g. gender or age); d) To toggle between the bar chart view and a box plot view, with expression values represented as a single point for each sample. Samples are split into the same groups whether displayed as a bar chart or box plot; e) To provide a color legend for the sample groups; f) To select categorical information that is to be overlaid at the bottom of the graph - for example, the user can display gender or smoking status in this manner; g) To provide a color legend for the categorical information overlaid at the bottom of the graph; h) To download the graph as a portable network graphics (png) image.

Measurements have no intrinsic utility in absence of contextual information. It is this contextual information that makes the results of a study or experiment interpretable. It is therefore important to capture, integrate and display information that will give users the ability to interpret data and gain new insights from it. We have organized this information under different tabs directly above the graphical display. The tabs can be hidden to make more room for displaying the data plots, or revealed by clicking on the blue “show info panel” button on the top right corner of the display. Information about the gene selected from the list on the left side of the display is available under the “Gene” tab. Information about the study is available under the “Study” tab. Rolling the mouse cursor over a bar chart feature while displaying the “Sample” tab lists any clinical, demographic, or laboratory information available for the selected sample. Finally, the “Downloads” tab allows advanced users to retrieve the original dataset for analysis outside this tool. It also provides all available sample annotation data for use alongside the expression data in third party analysis software. Other functionalities are provided under the “Tools” drop-down menu located in the top right corner of the user interface. Some of the notable functionalities available through this menu include: a) Annotations, which provides access to all the ancillary information about the study, samples and dataset organized across different tabs; b) Cross-project view; which provides the ability for a given gene to browse through all available studies; c) Copy link, which generates a mini-URL encapsulating information about the display settings in use and that can be saved and shared with others (clicking on the envelope icon on the toolbar inserts the URL in an email message via the local email client); d) Chart options; which gives user the option to customize chart labels.

Quality Control

Quality control checks can be performed on the datasets loaded on GXB, for example by examining concordance of the gender-specific expression of the XIST gene in those datasets for which gender information was available as metadata. The XIST gene is essential for imprinted and random X-chromosome inactivation and therefore, expression is expected to be high in female and low in male samples. Respective hyperlinks are found
<table>
<thead>
<tr>
<th>Title</th>
<th>Response to Virus/Vaccine</th>
<th>Cell type/Tissues</th>
<th>Number of samples</th>
<th>Selected marker for QC</th>
<th>Citation</th>
<th>GEO ID</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blood transcriptomes of human bacterial and influenza A pneumonia</td>
<td>Natural infection</td>
<td>Whole blood</td>
<td>190</td>
<td>Xis</td>
<td>GSE40012</td>
<td>9</td>
</tr>
<tr>
<td>Expression profiling of critically ill influenza and bacterial pneumonia patients, also influenza vaccination recipients.</td>
<td>Natural infection</td>
<td>Whole blood</td>
<td>81</td>
<td>ND*</td>
<td>GSE20346</td>
<td>14</td>
</tr>
<tr>
<td>FACs-sorted cells from Young Adults Vaccinated with Influenza TIV or LAIV Vaccines during 2008/09 Flu Season</td>
<td>Natural infection</td>
<td>Whole blood</td>
<td>84</td>
<td>ND*</td>
<td>GSE29618</td>
<td>16</td>
</tr>
<tr>
<td>Gene expression analysis in children with complex gastrointestinal infections</td>
<td>Natural infection</td>
<td>Whole blood</td>
<td>32</td>
<td>Xis</td>
<td>GSE2034</td>
<td>6</td>
</tr>
<tr>
<td>Gene expression analysis of Influenza vaccine response in Young and Old - Year 1</td>
<td>Experimental infection</td>
<td>PBMC</td>
<td>72</td>
<td>ND*</td>
<td>GSE6835</td>
<td>23</td>
</tr>
<tr>
<td>Gene expression analysis of Influenza vaccine response in Young and Old - Year 2</td>
<td>Natural infection</td>
<td>Whole blood</td>
<td>166</td>
<td>ND*</td>
<td>GSE1715</td>
<td>8</td>
</tr>
<tr>
<td>Genome-wide analysis of whole blood transcriptional response to Respiratory Synovial Virus (RSV), Influenza and Rhinovirus lower respiratory tract infection</td>
<td>Natural infection</td>
<td>Whole blood</td>
<td>36</td>
<td>Xis</td>
<td>GSE38900</td>
<td>8</td>
</tr>
<tr>
<td>Genome-wide analysis of whole blood transcriptional response to Respiratory Synovial Virus (RSV), Influenza and Rhinovirus lower respiratory tract infection</td>
<td>Natural infection</td>
<td>Whole blood</td>
<td>113</td>
<td>Xis</td>
<td>GSE38900</td>
<td>8</td>
</tr>
<tr>
<td>Genome-wide analysis of whole blood transcriptional response to Respiratory Synovial Virus (RSV), Influenza and Rhinovirus lower respiratory tract infection</td>
<td>Natural infection</td>
<td>Whole blood</td>
<td>205</td>
<td>Xis</td>
<td>GSE38900</td>
<td>8</td>
</tr>
<tr>
<td>Genome-wide analysis of whole blood transcriptional response to Respiratory Synovial Virus (RSV), Influenza and Rhinovirus lower respiratory tract infection</td>
<td>Natural infection</td>
<td>Whole blood</td>
<td>72</td>
<td>Xis</td>
<td>GSE38900</td>
<td>8</td>
</tr>
<tr>
<td>Genome-wide analysis of whole blood transcriptional response to Respiratory Synovial Virus (RSV), Influenza and Rhinovirus lower respiratory tract infection</td>
<td>Natural infection</td>
<td>Whole blood</td>
<td>166</td>
<td>Xis</td>
<td>GSE38900</td>
<td>8</td>
</tr>
<tr>
<td>Genome-wide analysis of whole blood transcriptional response to Respiratory Synovial Virus (RSV), Influenza and Rhinovirus lower respiratory tract infection</td>
<td>Natural infection</td>
<td>Whole blood</td>
<td>214</td>
<td>Xis</td>
<td>GSE49762</td>
<td>24</td>
</tr>
<tr>
<td>Genome-wide analysis of whole blood transcriptional response to Respiratory Synovial Virus (RSV), Influenza and Rhinovirus lower respiratory tract infection</td>
<td>Natural infection</td>
<td>Whole blood</td>
<td>30</td>
<td>Xis</td>
<td>GSE49762</td>
<td>24</td>
</tr>
</tbody>
</table>

*ND* indicates not determined.
<table>
<thead>
<tr>
<th>Selected marker for QC</th>
<th>GEO ID</th>
<th>Number of samples</th>
<th>Cell type/Tissues</th>
<th>Virus/Vaccine</th>
<th>Response to</th>
<th>Platforms used</th>
<th>Title</th>
</tr>
</thead>
<tbody>
<tr>
<td>GSE2938</td>
<td>Influenza</td>
<td>PBMC</td>
<td>Whole blood</td>
<td>Influenza</td>
<td>Vaccination</td>
<td>Affymetrix HuGene 1.0 ST</td>
<td>Host gene expression signatures of influenza A H1N1 and H3N2 virus infection in adults.</td>
</tr>
<tr>
<td>GSE4735</td>
<td>Influenza</td>
<td>PBMC</td>
<td>Whole blood</td>
<td>Influenza</td>
<td>Vaccination</td>
<td>Affymetrix HG-U133A</td>
<td>Host transcriptomic response to influenza and other acute respiratory viral infections – a prospective cohort study.</td>
</tr>
<tr>
<td>GSE5242</td>
<td>Influenza</td>
<td>PBMC</td>
<td>Whole blood</td>
<td>Influenza</td>
<td>Vaccination</td>
<td>Illumina HumanWG-6 v2</td>
<td>Host transcriptomic response to influenza and other acute respiratory viral infections – a prospective cohort study.</td>
</tr>
<tr>
<td>GSE6831</td>
<td>Influenza</td>
<td>PBMC</td>
<td>Whole blood</td>
<td>Influenza</td>
<td>Vaccination</td>
<td>Illumina HumanHT-12 v4</td>
<td>Host transcriptomic response to influenza and other acute respiratory viral infections – a prospective cohort study.</td>
</tr>
<tr>
<td>GSE2180</td>
<td>Influenza</td>
<td>PBMC</td>
<td>Whole blood</td>
<td>Influenza</td>
<td>Vaccination</td>
<td>Affymetrix HG-U133A</td>
<td>Host transcriptional response to influenza and other acute respiratory viral infections – a prospective cohort study.</td>
</tr>
<tr>
<td>GSE6960</td>
<td>Influenza</td>
<td>PBMC</td>
<td>Whole blood</td>
<td>Influenza</td>
<td>Vaccination</td>
<td>Illumina HumanHT-12 v4</td>
<td>Host transcriptional response to influenza and other acute respiratory viral infections – a prospective cohort study.</td>
</tr>
<tr>
<td>GSE6182</td>
<td>Influenza</td>
<td>PBMC</td>
<td>Whole blood</td>
<td>Influenza</td>
<td>Vaccination</td>
<td>HumanHT-12 v4</td>
<td>Host transcriptional response to influenza and other acute respiratory viral infections – a prospective cohort study.</td>
</tr>
<tr>
<td>GSE7481</td>
<td>Influenza</td>
<td>PBMC</td>
<td>Whole blood</td>
<td>Influenza</td>
<td>Vaccination</td>
<td>Affymetrix HG-U133A</td>
<td>Host transcriptional response to influenza and other acute respiratory viral infections – a prospective cohort study.</td>
</tr>
<tr>
<td>GSE4802</td>
<td>Influenza</td>
<td>PBMC</td>
<td>Whole blood</td>
<td>Influenza</td>
<td>Vaccination</td>
<td>Illumina HumanHT-12 v4</td>
<td>Host transcriptional response to influenza and other acute respiratory viral infections – a prospective cohort study.</td>
</tr>
<tr>
<td>GSE3420</td>
<td>Influenza</td>
<td>PBMC</td>
<td>Whole blood</td>
<td>Influenza</td>
<td>Vaccination</td>
<td>Illumina HumanHT-12 v4</td>
<td>Host transcriptional response to influenza and other acute respiratory viral infections – a prospective cohort study.</td>
</tr>
<tr>
<td>Title</td>
<td>Platforms used</td>
<td>Response to</td>
<td>Virus/Vaccine</td>
<td>Cell type/ Tissues</td>
<td>Number of samples</td>
<td>Selected marker for QC</td>
<td>Citation #</td>
</tr>
<tr>
<td>----------------------------------------------------------------------</td>
<td>----------------------</td>
<td>--------------------</td>
<td>---------------</td>
<td>-------------------</td>
<td>-------------------</td>
<td>-------------------------</td>
<td>------------</td>
</tr>
<tr>
<td>Transcriptomic profiling facilitates classification of response to influenza challenge</td>
<td>Illumina HumanHT-12 v4</td>
<td>Experimental infection</td>
<td>Influenza H3N2</td>
<td>Whole blood</td>
<td>88</td>
<td>ND*</td>
<td>31</td>
</tr>
<tr>
<td>Transcriptomic profiling in childhood H1N1/09 influenza reveals reduced expression of protein synthesis genes</td>
<td>Illumina HumanHT-12 v3</td>
<td>Natural infection</td>
<td>Influenza (H1N1), RSV</td>
<td>Whole blood</td>
<td>92</td>
<td>Xist</td>
<td>32</td>
</tr>
<tr>
<td>Trivalent Inactivated Influenza Vaccine (TIV) and Live Attenuated Influenza Vaccine (LAIV) Induce Different B cell and Transcriptional Responses in Children</td>
<td>Illumina HumanHT-12 v4</td>
<td>Vaccination</td>
<td>Influenza TIV, LAIV vaccines</td>
<td>Whole blood</td>
<td>140</td>
<td>ND*</td>
<td>30</td>
</tr>
<tr>
<td>Whole Blood Transcriptional Response to Pediatric Influenza Infection</td>
<td>Illumina HumanWG-6 v3</td>
<td>Natural infection</td>
<td>Influenza</td>
<td>Whole blood</td>
<td>31</td>
<td>Xist</td>
<td>8</td>
</tr>
</tbody>
</table>

*Gender information and/or Xist probe was not available. QC – quality control, ND – no data
Figure 2. Shown are XIST gene expression levels and gender information from the venipuncture validation set of GSE48762. Gene expression data were from whole blood of healthy adult volunteers before and after receiving either placebo (saline) injections, seasonal influenza (Fluzone) or pneumococcal (Pneumovax) vaccination.

in Table 1 allow you to visualize the XIST expression based on the gender information provided with the GEO submission. Figure 2 shows XIST gene expression in a representative dataset, along with gender information available that was recorded and made available in GEO.

Data availability
All datasets included in our curated collection are also available publicly via the NCBI GEO website: https://www.ncbi.nlm.nih.gov/gds/; and are referenced throughout the manuscript by their GEO accession numbers (e.g. GSE17763). Signal files and sample description files can also be downloaded from the GXB tool under the “downloads” tab.

Grant information
All the authors listed on this publication received support from the Qatar Foundation. Support for this project was provided by the Qatar National Research Fund [NPRP10-0205-170348].

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Acknowledgments
We would like to thank all the investigators who decided to make their datasets publicly available by depositing them in GEO.

References


Open Peer Review

Current Peer Review Status: ✔️ ✔️

Version 1

Reviewer Report 13 June 2019

https://doi.org/10.5256/f1000research.20283.r48849

© 2019 Tang B. This is an open access peer review report distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Benjamin M Tang
Department of Intensive Care Medicine, Nepean Hospital, Kingswood, NSW, Australia

This paper represents an extension of the authors' extensive work in this area - which aims to provide an accessible web-based platform for researchers to perform multi-cohort analysis of pooled transcriptomics data.

The Introduction outlines the scientific rationale to support the development of such a platform, which I mostly agree. There are a number of reasons why such a platform can assist researchers, including (1) help researchers to leverage on publicly available dataset to address each researcher's own research question, (2) obtaining a large sample size for analysis, which is otherwise not feasible by a single research group, (3) provide access to a diverse range of patient population and varying degree of disease severity, thereby generating more biological insights, which are not achievable by a single-centre study. For these reasons, the platform provided by these author represents an important contribution to the research community.

To further place this into perspective, one can estimate the likely cost (in time and money) to obtain the same analysis if such a platform is not available. To collect 31 transcriptomics datasets of viral respiratory infection (as outlined by this paper), a researcher need to perform an extensive literature review and database search. After all the datasets are collected, the researcher then need to bring in a bioinformatic team to perform a pooled analysis, which required advanced expertise due to the huge variability in technical platform, assay methods and normalization approach among different datasets. This bioinformatic work is extensive - if such work is outsourced to an external bioinformatician/company, the cost is in excessive of US$15,000 (our group has commissioned similar work in the past - hence our knowledge into the estimated cost). Here, the authors provides access to the analysis platform to the wider research community for free...an important step in democratising scientific data for the benefit of all researchers.

Is the rationale for creating the dataset(s) clearly described?
Yes

Are the protocols appropriate and is the work technically sound?
Yes

Are sufficient details of methods and materials provided to allow replication by others?
Yes

Are the datasets clearly presented in a useable and accessible format?
Yes

**Competing Interests:** No competing interests were disclosed.

**Reviewer Expertise:** Genomics, immunology, viral respiratory infection

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Reviewer Report 26 March 2019

https://doi.org/10.5256/f1000research.20283.r45689

© 2019 Moreira-Filho C. This is an open access peer review report distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Carlos Alberto Moreira-Filho
Department of Pediatrics, Faculty of Medicine, University of São Paulo (USP), São Paulo, Brazil

The paper by Bougarn et al. describes the use of an interactive web application, GXB (Speake et al.(2015)\(^1\)), for investigating the blood transcriptional response to viral respiratory tract infection and vaccination. A curated dataset collection was established for this purpose encompassing 31 datasets retrieved and selected from the NCBI Gene Expression Omnibus (GEO) public repository.

The rationale for creating the dataset collection and the methodology used in the paper are clearly described. The functionalities of the GXB tool are well explained to the reader, as well as the quality control procedures adopted by the authors. However, the GEO datasets do not always provide detailed information on virus genotypes. Distinct RSV genotypes induce very different PBMC transcriptional responses (see, for instance, Rodriguez-Fernandez et al. 2017\(^2\); and Vieira et al. 2019\(^3\)). The authors could insert a comment on this issue in their paper.

**References**

Journal of Infectious Diseases. 2018; 217 (1): 24-34 Publisher Full Text


Abstract | Publisher Full Text

Is the rationale for creating the dataset(s) clearly described?
Yes

Are the protocols appropriate and is the work technically sound?
Yes

Are sufficient details of methods and materials provided to allow replication by others?
Yes

Are the datasets clearly presented in a useable and accessible format?
Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: genomics

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

The benefits of publishing with F1000Research:

- Your article is published within days, with no editorial bias
- You can publish traditional articles, null/negative results, case reports, data notes and more
- The peer review process is transparent and collaborative
- Your article is indexed in PubMed after passing peer review
- Dedicated customer support at every stage

For pre-submission enquiries, contact research@f1000.com